

Materials for bone regeneration: Current types, bioactive mechanism and updated investigations

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Abstract

The exploration of bone repair materials has been continuously carried out. After several decades of development, bone repair materials have experienced from traditional ceramics, metals and polymers to modern smart hydrogels and multifunctional composite materials, etc. At the same time of designing and preparing the novel materials, more and more attention has been paid to their osteogenesis mechanism and immune response. The development of next generation of bone repair materials requires the comprehensive consideration of a combination of many related factors. This article reviews the research status and the progress of bone repair materials from the three aspects, current most-used types, bioactive mechanism and updated related investigations. Finally, several research points that are crucial for the further development of bone repairs are proposed in the conclusion and perspectives part.

Introduction

Bone defects beyond critical-size due to the injury, tumor or osteomyelitis, which affect revascularization and tissue regeneration directly, may not be repaired without external interventions. Nearly 4 million bone fractures happened in US in 2013. Due to the ageing population, by 2025, the global costs of osteoporotic fracture are expected to increase by 25%.¹ Another statistics states that in 2015, around 7 million people in America underwent total hip or knee replacement.² Bone repair materials have huge market prospect.

Among all bone substitutions, autologous bone grafts are still considered as the *gold standard*, but the size and source limit its application, as well as the risk of infection and ongoing unendurable pain after a second surgery at the donor site. Allograft is an alternative to autografts, but it may be immunogenic or have the risk of viral transmission, which can lead to a high failure rate. On this occasion, available natural bone grafts are far from satisfying the clinical requirements.

Based on these facts, ceramics, synthetic or natural polymer materials, metals etc. are commonly used as bone repair materials with large supplement and less or free immunogenicity. They may not only provide structural support, but also have a promoting effect on bone regeneration, which is effective to repair large bone defects. It was suggested that bone repair materials should have the ability to promote osteogenic differentiation of hMSCS or the proliferation of osteoblasts,3 and ideal scaffolds for bone tissue engineering needed to have interconnected pores, appropriate porosity as well as suitable mechanical properties. Some studies emphasized on the ability to induce and support angiogenesis.⁴ A suitable pore size was necessary for blood vessel growth, transport of nutrients and metabolic waste.

This review outlines the current types of bone repair materials, from the ceramic, metal and polymer materials, carbon materials, to the smart hydrogels, multifunctional composite materials and natural biological material *etc.*, bioactive mechanisms involved and recent investigations, including functional improvements and immunological researches. Finally, this review analyses the future developments of bone repair materials.

Current materials and their applications

At present, there are many types of bone repair materials, such as bioceramics, metals, polymers and composites. Besides, carbon materials are also widely used. Currently, smart hydrogels, metal ions doped composites and natural biomaterials *etc.* have been well developed. Some of these have made remarkable achievements in animal models or clinical researches.

Conventional materials

Bioceramics and bioactive glasses

Calcium phosphates (CAPs) and bioactive glasses are commonly used as bone repair materials. CAPs, including hydroxyapatite (HA), tricalcium phosphate (TCP) and Correspondence: Xiaoming Li, Key Laboratory for Biomechanics and Mechanobiology of Ministry of Education, School of Biological Science and Medical Engineering, Beihang University, 37 Xueyuan Rd, Beijing 100083, China.

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biphasic calcium phosphate (BCP), have good biocompatibility and osteogenic activity.⁵ It had been shown that the CAPs could form strong chemistry bonds with surrounding bone and promote osseointegration.⁶⁻⁸

HA is similar with the main component of natural bone,⁹ and can bond with natural bones in body. With good osteogenic activity, it had been used as bone repair materials before century 21.¹⁰ The surfaces of HA can not only provide nucleating sites for the precipitation of apatite crystals in culture medium,^{11,12} but also accelerate apatite growth.¹³ A study showed that HA could be slowly degraded by simple dissolution or by osteoclastic bone remodeling effect.¹⁴ However, with low breaking strength and high brittleness, it may be unsuitable to be directly used in large-scale bone defect repair. Fortunately, with better bioactivity and higher mechanical strength, nano-HA may be an ideal bone repair material.¹⁵ For example, Kubasiewicz *et al.*¹⁶ created 5 mm calvaria bone defects in rats and used nano-HA to repair. Histological analysis and micro-CT evaluation showed that 34.2-44.4% new bone was formed after 8 weeks, whereas only 13.0% in empty control group.

β-TCP, a good osteoconductive material, was reported to have similar effects to inorganic bovine bone (Bio-Oss, Geistlich Pharma, Wolhusen, Switzerland) in criticalsize defect repair of rats.¹⁷ For patients with tibial plateau fractures, a long-term study showed that new bone was formed around β-TCP with good functional recovery, which meant that β-TCP was an effective material in tibial plateau fracture treatments.¹⁸

BCP ceramic, a mixture of HA and TCP, has been widely used in clinical. It is reported that the BCP might be a feasible alternative to autografts.¹⁹⁻²³ In the surgical management of scoliosis, the correction was maintained similarly in BCP and autografts groups, but BCP groups had lower blood loss and free postoperative local complications.20 And in another study, BCP could achieve equivalent fusion rates to autograft in rabbit lumbar arthrodesis models.²¹ In addition, Fellah et al.23 compared the osteogenicity of BCP granules with autograft, the results of which showed that the BCP had better osteogenic property and stability in critical-sized bone defect repair.

Bioactive glasses are biodegradable in physiological conditions. They can release ions, such as Na⁺ and Ca²⁺, to give rise to new bone regeneration with mechanically strong bond to the surfaces of implanted.²⁴ They have osteoconductivity, and can bond firmly both with natural bones and soft tissues. However, high brittleness limits their applications to repair load-bearing bones.²⁵ So, it is pressing to optimize bioactive glasses by innovative scaffold design and processing.

Metals and their alloys

As early as last century, due to high mechanical strength, metals and their alloys have been widely used for load-bearing applications in bone repair, including bone pins, plates, screws, knee or hip prostheses and dental implants, *etc.*²⁶ Alloys, which generally perform better than pure metals, are more commonly used. Stainless steel, Co-based alloys and Ti-based alloys are the three main types for bone repair.

With low cost and good processibility, stainless steel, such as 316L stainless steel,

today. However, many studies had shown that the stainless steel often underwent corrosion failure in the body²⁷ or in simulated body fluids,²⁸ especially with the existence of Cl-.29 As a matter of fact, stainless steels are mostly used for temporary implant devices.26 Co-based alloys, like Co-Cr and Co-Ni, have been made for hip joint prostheses or other devices with finestructures.²⁶ They display high fatigue strength and wear resistance. It was reported that Co-based alloys had better corrosion resistance compared to others.30 However, it was reported that in long period implantations, a high amount of released Cr and Ni ions might lead to negative toxic effects31 or allergic responses,32 etc. Moreover, Ti is a suitable option for bone repair with superior biocompatibility, low density (4.5g/cm³), good corrosion resistance, low elastic modulus, and non-toxic even in large doses.29 Researches showed that Ti and Ti-based alloys could make good physical connections with the host bone³⁰ and had been widely used in clinical, such as total hip replacements and craniofacial implants.²⁶ However, there are some common problems of metals and their alloys, such as the stress shielding effect due to the mismatch between elastic modulus of metal implants and that of natural bone, corrosion and fatigue fractures, inflammatory reactions, and allergic reactions caused by the wear debris or metal ions. One promising strategy to achieve suitable elastic modulus is to prepare porous metallic implants. It is also needed to control the corrosion rate to avoid metal-related toxicity by precise modification and control of micro- and nano-structures.

is still widely used for implant devices

Polymers

Polymers can be divided into natural and synthetic ones. Natural materials involve collagen, silk fibroin (SF), chitin (or chitosan), hyaluronic acid, sodium alginate, *etc.* Synthetics mainly include polylactic acid (PLA), Polycaprolactone (PCL) and polylactic acid-glycolic acid copolymer (PLGA) and so on.

Collagen is the main organic component in natural bone and serves as the template for mineralization with specific structures and bioactivities. However, pure collagen is unsuitable for direct applications due to its poor mechanical strength and high degradation rate. Fortunately, reports showed that it could encapsulate the MSCs as injectable microspheres for bone repair,³³ or combine with bioceramics or other polymers to further increase the mechanical and biological properties.³⁴ pagepress

obtained from spiders or silkworms silks.35 With satisfactory flexibility, extensibility and tensile strength, it can be molded into different kinds of forms.36 It can offer active sites to aid mineralization or bond with bioactive molecules to promote bone regeneration. It was reported that the electrospun silk fibroin scaffolds could facilitate the new bone formation in rat calvaria defect models more significantly than PLA scaffolds.37 Moreover, the degradation rate of silk fibroin could be controllable, ranging from weeks to a year.38,39 So, SF is expected to be widely applied to repair bone in the phase of human trials soon.40 With varied mechanical properties and processability, PLA is widely used for bone repair in clinical.41 Up to now, there are many types of biodegradable PLA devices, such as screws, plates and pins, which, unlike alloys, can not only withstand bear-loading and transfer stress to the damaged area to avoid stress shielding effect over a period of time, but also avoid a second surgical procedure to remove. However, they have some weaknesses, such as poor hydrophilicity and excessively rapid degradation rate, etc. Moreover, the accumulation of the degradation product, lactic acid, in vivo might lead to a too low local pH, which could cause inflammatory reaction42 or even osteolysis.43,44 Therefore, to neutralize the acidity or to modify properties, PLA was often composite with other fillers, such as HA, chitosan.45

In short, both natural and synthetic materials have their own advantages and disadvantages. For example, on the one hand, most natural ones have low immunogenicity, no cytotoxicity, good biocompatibility and bioactivity. Some of those have high mechanical properties, osteoconductivity or osteoinductivity. Besides, the structures of some natural materials are similar to those of the natural bone ECM. However, some natural materials are difficult to obtain or process,46 and sometimes the performance may vary with batches. Furthermore, chemical or enzymatic treatment may damage their functionality or even result in inactivation. On the other hand, synthetics have well-adjustable mechanical properties and outstanding processability, which can be easily modified into various shapes or to mimic natural matrices. But some have poor biocompatibility, too rapid or slow degradation rate, less integration with the surrounding tissue.

Composites

A single material often has inevitable shortcomings, so people turn their attention to the composites, which may combine the positive functionalities of each component



and compensate the disadvantages of the individual or even have new properties that are not available in either material used alone.⁴⁷

Polymers, ceramics and metals can be used as matrixes or reinforcements for bone repair composites to acquire better properties. For example, Li *et al.*⁴⁸ prepared nano-HA/collagen/PLLA composites reinforced by chitin fibers, which had higher compressive strength than nano-HA and were similar to natural bone in components with the existence of the chitin fibers. The further study *in vivo* showed that the composites could repair 40 mm goat shank bone defect successfully in 15 weeks, which is firstly reported to show that the use of biomaterials repaired completely segmental bone defects larger than 30 mm.

In addition, Chen et al.49 prepared and investigated the collagen-SF/HA nanocomposites, which had suitable structural properties, good biocompatibility in vitro and an enhanced elastic modulus with the presence of SF in comparison to collagen/HA composites. In another study, Zhang et al. developed small intestinal submucosa/polymethyl methacrylate (SIS/PMMA) composites for vertebral repair in rats. With a higher porosity, decreased stiffness and greater osteoinductivity, they could enhance the adhesion, proliferation and osteogenic differentiation of MC3T3-E1 cells and BMSCs compared to PMMA. And further study indicated that these composites can greatly enhanced osteointegration and bone regeneration in vertebral defect models.50 In this system, PMMA serves as a basic material with stiffness and formability, and the bioactive SIS enhanced the biological performance. Moreover, it was reported that the PLGA/TCP porous scaffold with Mg ions incorporated had good bioactivity and enhanced mechanical properties. As the released of Mg ions in this system raised the activity of osteoblasts, this scaffold could promote more new bone formation than PLGA/TCP scaffold in 15 mm radius defects of rabbits.51

Carbon nanomaterials

With excellent mechanical properties and biocompatibility, carbon materials, especially carbon nanotubes (CNTS) and grapheme, have attracted more and more attention.

CNTs are cylindrical nanostructures and have extraordinary properties, like good tensile strength, high aspect ratio⁵² and good biocompatibility *in vivo*.^{53,54} Generally, they are often used as reinforcing nanofillers in bone repair composites. It was reported that the stiffness of CNT-ploymer composites could be increased by 7 times.⁵² Another research showed that the HA-CNTs composite had considerable improvement in fracture toughness and flexural strength over pure HA, and the toughness of this composite was close to that of human bone. Further study proved that it could enhance new bone formation with strong interactions and high interfacial strength with the host bone in rabbit models.55 Though some studies showed that carbon nanotubes were poor in hydrophilicity and easy to be agglomerated, they could be functionalized⁵⁶ or modified on the surface to remedy the weaknesses. For example, Venkatesan et al.57 fabricated chitosan-MWNTs composites for bone repair. This bicomponent system had increased porosity and could enhance cells proliferation and ALP expression than chitosan scaffold. It was also demonstrated that after carboxyl-functionalization, the MWNTs were uniformly distributed with the chemical interactions formed with chitosan. Besides, it had been reported that the specific treated multiwalled carbon nanotubes (MWNTs) could absorb bone-formed related proteins to promote stem cells differentiate into osteoblasts.58,59

Another promising carbon nanomaterial, grapheme, is an allotrope of carbon with a single atomic layer of six-atom rings network, which has many unusual properties, such as large specific surface area,⁶⁰ distinguished electrical conductivity and mechanical properties.⁶¹ Many studies showed that grapheme had potential good biological property and could improve the adhesion, proliferation and differentiation of osteoblasts,⁶² which indicated that it had marvelous potential in bone repair. However, there are still some shortcomings of graphene, such as the long-term toxivcity and poor biodegradability.⁶³ So, its applications in bone repair still remain studies *in vitro* or animal experiments stages.

Smart hydrogels

Although the concept of hydrogels appeared at the end of the 19th century, the word smart was not introduced until 1948 by Kuhn.64 The smart hydrogels can not only present specific structures and functions by one or more stimuli, like temperatures, pH, electric, magnetic fields, light and biomolecule, but also be customized for different targets, such as chronic inflammation treatment, cancer therapy and bone repair.65 Nowadays, smart hydrogels have become research hotspots in bone repair. The number of publications in the Science Direct database about smart hydrogels and smart hydrogels for bone repair since 1998 has had a booming growth (Figure 1).

It has been showed that the release rate of the bioactive proteins, cells or drugs loaded in smart hydrogels with threedimensional structures could be controlled by different external stimuli,^{66,67} thereby providing potentials as intelligent drug delivery systems in a controlled manner for bone repair applications. Papathanasiou *et al.*⁶⁸ prepared the programmable and responsive silica-based hydrogels, with bisphosphonates (BPs) loaded, for osteoporosis therapy. The release rate of BPs could be

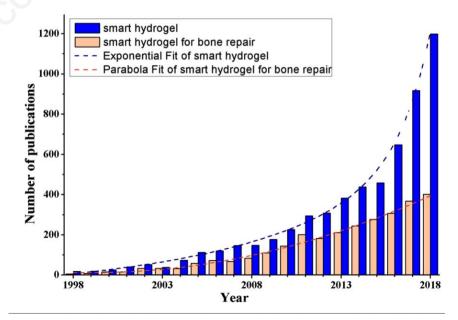
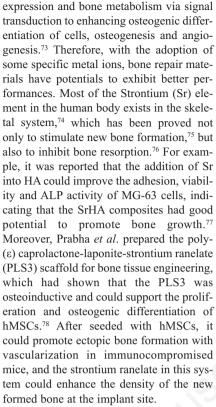


Figure 1. The number of publications about smart hydrogels or smart hydrogels for bone repair in Science Direct database per year. (Compiled from a literature search in Science Direct database in December 2018).

fine-tuned by several factors, like cations, pH, temperature and side-chains of BPs, to meet the patient's requirements. For example, the change of pH could reinforce or weaken the binding strength of hydrogen bonds in the hydrogels to affect the gelation to control the release rate of BPs. Moreover, injectable smart hydrogels can perfectly fit defects of different sizes and irregular shapes with homogenous distribution, minimal invasion and low risk of infection. They can perform good integrations with host bone and enhance new bone formation. For example, one kind of reported injectable acellular bone matrix/PEG-PCL-PEG hydrogels were bioactive and thermosensitive, which could be molded into the shape of the defects perfectly and gel in a few minutes at body temperature in cranial defects of rabbits. The X-ray examination, micro-CT images and histological analysis all proved that these hydrogels could be effective in cranial defects repair, and the density of new formed bone was approximate to that of the host cranial bone after 20 weeks.69 Furthermore, it is well known that dynamically tunable cell culture platforms are similar to natural cell microenvironments. Based on their stimuliresponsiveness to pH, temperature, radiation, etc., smart hydrogels can provide various dynamic chemical, physical, and mechano-structural cues to the adhered cells in the platforms, where the fundamental biological processes can be deeply understood, and the interactions between implants and cells can be well simulated.70 The smart hydrogels in these platforms can present different mechanical properties to simulate the in vivo ECM to control cellular processes. For example, Yang et al. developed a phototunable PEG hydrogels with tunable stiffness to stimulate the highly dynamic ECM, which could be used to investigate the response of hMSCs to the stiffness of the substrate and whether the MSCs had the memory of the previous mechanical signals.71 With significant advantages, smart hydrogels are commonly used in bone repair and delivery system.72 But, it is still needed to consider about controlled biodegradability and the maintenance of mechanical properties after implantation.

Metal ions

Natural bone microenvironment contains a variety of bioactive metal ions, such as Strontium, Zinc and Magnesium ions, which often exist in many surrounding macromolecules, like enzymes and nucleic acids, *etc.* It has been reported that some metal ions could induce bone related gene



Magnesium (Mg) ions are bio-safe both *in vitro* and *in vivo*, and can be promptly diluted and excreted.⁷⁹ They exist in natural bone and can stimulate bones growth⁸⁰⁻⁸² or remodeling.⁸³ Mg ions perform as cofactors in various bone-related enzymatic reactions, such as protein and nucleic acid synthesis, energy metabolism and vitamin D metabolism.⁸⁴ As mentioned above,⁵¹ PLGA/TCP porous scaffold incorporated with Mg ions could promote osteogenesis. Another study showed that the presence of Mg in TCP scaffolds could facilitate angiogenesis in femoral defect of rat models.⁸⁵

Zinc (Zn) ions are also necessary in protein synthesis, cell proliferation and DNA synthesis, which can stimulate bone formation and inhibit bone resorption. For example, Bhattacharjee et al.86 doped Zn ions in HA to improve its bioactivity. After the implantation in femur bone of rabbits for two months, the percentage of new bone formation in Zn-HA group was significantly higher than that of pure HA group. Histological studies showed that welldevelopment of Haversian system and Volkmann's canal was formed in the regenerated bone. Moreover, mechanical pushout testing demonstrated stronger interfacial strength between the Zn-doped implants and host bone. Another study showed that in Zn-doped TCP cement, the low concentration of Zn²⁺ could promote the proliferation and ALP activity of rat MSCs to enhance bone formation.87 Furthermore, the Zn2+ could offer anti-bacterial property to TCP to



prevent clinical infections. However, high concentration of Zn^{2+} might restrict cell growth,^{88,89} so it is necessary to control the amount of Zn^{2+} in the composites and their release rate.

In fact, the current use of metal ions in bone repair materials mostly stays in the stage of animal experiments only with few clinical results, so further studies on the doping methods and precise release of metal ions are necessary.

Natural biomaterials

With specific advantages, such as low immunogenicity and similar structure to that in the human body, natural biomaterials for bone repair have attracted much attention. Among all, those derived from aquatic organisms, like corals and fish collagens, with a wide range of source, good bioactivity and osteogenic activity, have good potential for bone repair. As early as the 1980s, corals had been investigated and used as bone repair materials.90 Nowadays, there are some effective commercially available products in clinical, like Pro Osteon[™] and Biocoral[®].⁹¹ The structures of corals were networks with interconnected channels and pores, which were highly similar to those of cancellous bones and could allow for the ingrowth of capillaries.92 Besides, it had been proved that hMSCs and osteoblasts could adhere, proliferate or differentiate on the coral scaffolds with high DNA content and ALP activity.93,94 Nevertheless, in some other studies, corals had relatively low resorption and fusion rate in cervical arthrodesis,95-97 and poor bone ingrowth at the defects of iliac crest,98 which indicated that the experimental results were inconsistent and more in-depth investigations into their long-term performances were needed. Moreover, the overexploitation and environmental changes had put them in an endangered position.94 Meanwhile, corals might contain some toxic elements due to the environment pollution.99 So, it is necessary to launch better protection measures and detailed examinations into corals before considering the application to bone repair, or try to cultivate artificial corals. Due to its wide range of sources, low cost and minimal disease transmission,100 fish collagen is considered as an appropriate alternative to mammal collagen for bone repair. It was reported that the bioactive tilapia collagen could increase the expression of ALP, osteocalcin and osteopontin in pre-osteoblasts in vitro.101 Moreover, Matsumoto et al. demonstrated that the flexible and soft tilapia collagen could induce the adhesion and the early osteoblastic differentiation of hMSCs better than the hard porcine collagen. And the



aligned tilapia collagen fibrils could induce cell polarization and facilitate osteogenesis while the randomly arranged porcine collagen didn't have similar functions.¹⁰² However, it was showed that fish collagen might be allergens for some people.¹⁰³ Therefore, more hyposensitization and detailed safety tests of fish collagen should be taken into consideration.

Bioactive mechanism of the materials

Generally, there had been many studies on the approaches for designing the bone repair materials and the expression of bonerelated gene or proteins, but the intrinsic bioactive mechanisms were often ignored.104,105 Fortunately, more and more recent studies have begun to focus on their bioactive mechanisms. Nowadays, it has been shown that bone repair materials exhibit their functions by affecting signaling pathways of osteogenic-related cells, such as Notch, Wnt and BMP¹⁰⁶ (Figure 2). For example, Jung et al. reported that the silk fibroins could up-regulate the expression of ALP and Runx2 mRNA by inhibiting Notch signaling pathway of bone marrow cells to regulate osteogenesis.107 Moreover, studies showed that, by significantly up-regulating the activity of BMP2108 and Wnt109 signaling pathways of MSCs, the β-TCP could stimulate the bone regeneration. In another study, Zhang et al, showed that Mg2+ could increase the expression level of Runx2 and ALP to significantly improve the osteogenic activity via TRPM7/PI3K signaling pathway of human osteoblast cells.110 Besides, some composites have been shown to promote bone repair with the similar mechanism. For example, Liu et al. had demonstrated that the nano-HA/chitosan scaffold could activate the integrin-BMP/Smad signaling pathway to up-regulate the mRNA expression level of the BMP-2/4, Runx2 and ALP to induce the osteogenic differentiation of mBMSCs.111 Meanwhile, the physical and chemical properties of materials have effects on signaling pathways. For example, studies had proven that the surface micro/nanotopography of materials could influence the Hedgehog-Gli1,112 FAK-ERK1/2,113 ILK/ERK1/2 and ILK/p38114 signaling pathways to regulate cell fates. In addition, it had been found that the increasing matrix stiffness could activate the Wnt signaling pathway to promote the proliferation and osteogenic differentiation of dental pulp stem cells,¹¹⁵ or could up-regulate the expression of macrophage migration

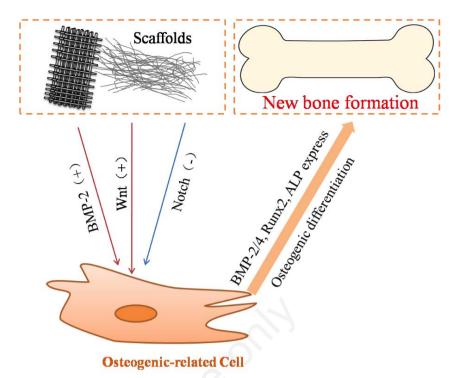


Figure 2. The bone repair materials can promote cell proliferation and differentiation through affecting the Notch, Wnt and BMP-2 signaling pathways to stimulate the expression of BMP-2/4, Runx2 or ALP, etc. in osteogenic-related cells to promote new bone formation.

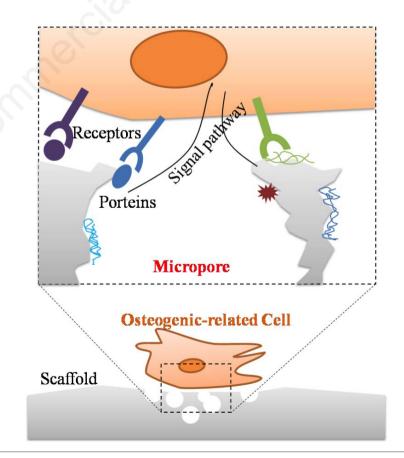


Figure 3. The porous scaffold can adsorb different proteins *in vitro* to promote cell adhesion and proliferation (the walls of pores are showed in grey).

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inhibitory factor (MMIF) to activate the AKT/YAP/RUNX2 signaling pathway to facilitate osteogenic differentiation of hMSCs.116 Furthermore, the porous nanomaterials could provide adsorption sites to gather and accumulate a large amount of bone-related proteins from the culture medium to promote cell adhesion and proliferation (Figure 3).117-119 Moreover, the recruitment of endogenous MSCs in vivo is critical to bone regeneration. It had been hypothesized that MSCs might migrate to the targeted sites due to the participation of chemokines, growth factors, or other chemical or physical cues from bone repair materials. For example, it was reported that the bFGF-loaded acellular dermal matrix (ADM) could recruit a plenty of MSCs to the implant sites and promote their proliferation to a sufficient amount to facilitate bone regeneration.120 In another study, Shih et al. showed that the PEGDA-co-A6ACA hydrogels could promote the osteogenic differentiation of MSCs, ES and even iPSCs without the addition of growth factors. After the subcutaneous implantation in the spine of rats, it could recruit endogenous progenitor cells for bone formation by the simulation of the bone environment to assist spinal fusion.121,122 In addition, Song et al. implanted the osteoinductive microporous BCP in dogs' dorsal muscles and observed that the osteoprogenitor cells and BMSCs were homed from the bone marrow to the implant site through blood circulation to induce ectopic bone formation.123

Updated investigations of bone repair materials

Nowadays, bone repair materials have been continually optimized to improve their biocompatibility, bioactivity and osteogenic capability, and to overcome the shortcomings. Furthermore, the immunoregulation effect of materials was also underlined in some studies.

Function improvement

Recently, with the development of manufacturing technology and the in-depth researches, more and more multifunctional bone repair materials have appeared. On the one hand, these materials not only have the function of the conventional materials, but only have suitable degradation rate, antibacterial property and the function to promote vascularization, *etc.* On the other hand, these conventional functions, bioactivity and osteogenesis ability, *etc.* had been further improved.

Multi-function

Much attention has been paid to the multifunctionality of bone repair materials to obtain better repair efficiency. For example, in a recent study of Li et al., with the reinforcement of silica nanoparticles (SNs), the biodegradable and bioactive poly (citrate-siloxane) had the enhanced photoluminescent capacity for imaging application. The further *in vitro* experiment showed that the osteoblasts (MC3T3-E1) had improved attachment and proliferation on this material. Therefore, it was indicated that this material had the potential to realize simultaneous bone regeneration and fluorescence imaging.¹²⁴ In another study, with Co ions incorporated, the bioactive glass-collagenglycosaminoglycan scaffold could support osteogenesis and promote the expression of VEGF, indicating that this scaffold has both pro-angiogenic and pro-osteogenic capabilities for bone repair.125

Bioactivity improvement

In order to further improve the bioactivity and osteogenesis ability of materials, bioactive molecules, like peptides and growth factors, have been adopted into the bone repair materials.

Recently, osteogenesis-related polypeptides have been widely used in bone repair materials due to their outstanding osteogenic ability, low cost, stability and minimal tumor-related side effects.126 Generally, these polypeptides can be bonded to the surface of materials by either noncovalent or covalent bonds. But the covalent combination can effectively increase the interactions between the polypeptides and materials, prolong the duration of polypeptides action and make the functionalized surfaces more stable than the noncovalent one.127 It was reported that, many polypeptides could be added in bone repair materials to mimic native bone ECM. Among them, Arg-Gly-Asp (RGD) polypeptide, which could be found in fibronectin and other molecules in ECM, could be coated on bone repair materials to enhance the adhesion and differentiation of osteoprogenitor cells.128

Loading growth factors on bone repair materials is another common method to improve the osteogenesis ability. For example, Zhao *et al.* immobilized BMP-2 on the surfaces of PLGA/HA fibrous scaffolds to increase the expression of osteogenesisrelated genes and up-regulate the activity of ALP *in vitro*.¹²⁹ In another study, Kuttappan *et al.* stated that the sustained local delivery of multiple growth factors might be a potential approach for bone repair.¹³⁰ They developed PLLA-nanoHA-gelatin composite scaffolds with dual growth factor (VEGF-



BMP-2 or FGF-2-BMP-2) loaded, by dissolving these growth factors with specific ratio in phosphate buffered saline (PBS, pH 7.4). Further in vivo study showed that, during 24 h, about 23 % of BMP-2, 42% of FGF-2 and 85% of VEGF were released steadily. VEGF could be released totally within 7 days while BMP-2 and FGF-2 could be released till 20 days. Increased new vasculature in the scaffolds and new bone growing from the peripheral region towards the mid region of the defect were observed in calvarial defects of rats, the results of which demonstrated that these scaffolds could effectively promote vascularisation and bone regeneration.

Immune response

As foreign bodies, bone repair materials can inevitably cause immune responses after the implantation. The interactions between materials and the surrounding immune environment should be considered, as well as the assessment of the osteogenic capacity of bone repair materials. For example, in a research of Chen et al., hMSCs and macrophages were cultured alone or together on cobalt incorporated β-tricalcium phosphate (CCP). When hMSCs were cultured alone, the CCP had the effect to promote their osteogenic differentiation. But this effect was reduced with the presence of macrophages. After the implantation in femur of rats, CCP was coupled with fibrous encapsulation rather than promoted new bone regeneration. The research showed that the CCP could shift macrophages to an M1 phenotype and cause enhanced inflammation. Therefore, the excessive inflammation impeded the osteogenic differentiation.131

Furthermore, other studies reported that macrophages played key roles in the recruitment and differentiation of MSCs. It was a common understanding that the coordinated cross-talk between immunocyte and bone related cells, especially macrophages and MSCs, could contribute to successful bone healing.¹³²

Moreover, it has been shown that materials with different surface properties might induce diverse types or extent of immune responses.¹³³ For example, the surface roughness and hydrophilicity could influence the interaction between immune cells and materials. Hotchkiss *et al.* showed that, by increasing the roughness and surface wettability, titanium implant could polarize the adaptive immune response towards a Th2, pro-wound healing phenotype, to lead to a faster resolution of inflammation, by which, this implant could recruit more MSCs to the implant site with the presence of macrophages.¹³⁴



Conclusions

Although rapid developments on bone repair materials, there has not been a material that fully meets the ideal requirements, especially for large bone defect repair. Most materials remain in the *in vitro* or animal experiments. With the advancement of science and the improvement of people's living standards, the performance of bone repair materials is undoubtedly facing higher requirements and challenges.

Firstly, it is needed to produce bone repair materials that have multiple functions or can simulate biological processes. Materials are expected to accurately assist the natural healing process via additional functions. The treatment plan and the implanted material should be different for the bone defects caused by different reasons. For example, i) for trauma and infection, the materials should have antibacterial and anti-infective properties by adding chitosan, silver ions or loading drugs; ii) For bone tumor, after excision of the tumor, the cancer cells left at the edge of the defects will multiply and cause local recurrence. The implanted bone repair material should also have the ability to inactivate cancer cells to prevent recurrence while promoting bone regeneration; iii) For fractures due to osteoporosis, it is better that the implanted material can not only promote the homing of osteoblasts and stimulate bone regeneration in situ, but also release Ca2+ to increase the calcium content and bone mineral density of host bone, thereby reducing the rate of fracture recurrence.

Secondly, by structural adjustment, bone repair materials can be effectively combined with bioactive molecules and achieve stable and precise release for a sufficient period of time. It is a viable way to wrap or physically or chemically bind porous microspheres with bioactive molecules inside with bone repair materials. Another promising method for precise release is preparing multilayered scaffolds with different types or dose of growth factor. Besides, the materials may have the structure to adsorb specific growth factors. After implanted in the body, they can accumulate growth factors, and then release them gradually. Moreover, efforts to design materials with specific structure to stimulate related cells to express osteogenic proteins, and then adsorb the proteins on their surface to further bring positive effects to other attached cells, have been another promising research direction for bone repair materials.

Thirdly, in the wake of development in high technology, the approaches of process-

ing and preparing materials are more diversified. Advance techniques, such as 3D printing, and even the emerging 4D printing,¹³⁵ can manufacture materials with more complex structures to satisfy precious requirements. 3D CT scan image can capture the sophisticated details of the defects accurately. By the conversion of 3D CT scan image into mathematical model, biomimetic customized implants can be manufactured accurately through 3D printing. These techniques make it promising to realize the personalized medicine.

Fourthly, after implanted, materials are in a dynamic biomechanical environment instead of static. However, most of the current in vitro studies often ignore the influence of biomechanics. For example, although many bone bioreactors have been developed in vitro, they still have limitations and cannot fully simulate the complex environment of living organ. In many cases, the rates of material degradation in vivo and in vitro are different because of biomechanical environment. Thus, more attention should be paid on the effect of mechanics during the investigations into the degradation of materials in simulated body fluids or co-culture with cells. The in vivo mechanical environment can be simulated by adding a near physiological load. It is necessary to systematically study the relationship between material structures, mechanical properties and degradation in biomechanical environment to optimize bone repair materials. Moreover, at present, most of the results obtained in animal experiments may not be repeated in clinical.¹³⁶ On the one hand, a material should be tested from small animals to large ones systematically. Generally, small animals, such as rats and rabbits, have fast bone turnover rates and low cost, which can be used to assess the performances of materials initially. The advantage of large animals, such as goats and pigs, is that their physiology and biomechanical properties are closer to those of human being. So, experiments from small to large animals can help to find the rules of the repair process. Based on the comparison of the results obtained from small animals with those from large animals, the performances in clinical can be predicted. On the other hand, at present, there is hardly clinical standard to evaluate the degree of bone regeneration in animal experiments and clinical. So, a well recognized material should be selected first. Then, systematical and comprehensive investigations on this material both in animal models and in clinical are performed. Based on the results, an industry standard can be established to evaluate performances of other materials in the subsequent studies.

Fifthly, it is well know that vascularization, which is essential for the transport of nutrients and the recruitment of cells of the implant materials, is a prerequisite for bone healing,¹³⁷ especially for large bone defect repair. Although there are a lot of related researches, some of which took pore size,138 vascular-related growth factors and cell coculture,139 as well as hypoxia-induced signaling140 into account. Cell sheet technology, pre-patterning and 3D printing strategies are also worth trying,141 deeper understanding and study of the interactions between materials and endotheliocyte or related cells are necessary. Furthermore, it may be an effective solution to prepare bone repair containing some specific tubes loaded with vascular-associated growth factors.

Finally, bone repair materials with specific immunomodulatory functions should be another important future development direction.131 Immune cells play important roles in regulating skeletal dynamics. After materials are implanted, the first cells that they encounter are immune types. The regulation of the immune cell response by the material determines to some content whether bone regeneration can be promoted. At present, it is necessary to launch a large number of systematical investigations into the effects of the structure, composition and surface properties, etc. of bone repair materials on the differentiation of immune cells.

In short, the preparation of suitable bone repair materials is required to consider the multiple effects of various factors. Every factor is crucial and complements each other. It is necessary to advance longterm, cross-discipline collaborations. Further studies on the structure and properties of the materials, the mechanisms of angiogenesis, immune response and bone regeneration are imperative. With the above comprehensive developments, it should be much easier to achieve the satisfactory bone repair materials to meet the requirements of different patients for precision medicine.

References

- 1. Hernlund E, Svedbom A, Ivergård M, et al. Osteoporosis in the European Union: medical management, epidemiology and economic burden. Archiv Osteoporosis 2013;8:1-136.
- 2. Maradit Kremers H, Larson DR, Crowson CS, et al. Prevalence of Total Hip and Knee Replacement in the United States. J Bone Joint Surg Am 2015;97:1386-97.
- 3. Yunus Basha R, Sampath Kumar TS, Doble M. Design of biocomposite



materials for bone tissue regeneration. Mater Sci Engin C, Mater Biol Appl 2015;57:452-63.

Review

- Saran U, Gemini Piperni S, Chatterjee S. Role of angiogenesis in bone repair. Archiv Biochem Biophys 2014;561: 109-17.
- 5. Bouler JM, Pilet P, Gauthier O, Verron E. Biphasic calcium phosphate ceramics for bone reconstruction: A review of biological response. Acta Biomater 2017;53:1-12.
- Denry I, Kuhn LT. Design and characterization of calcium phosphate ceramic scaffolds for bone tissue engineering. Dental materials: official publication of the Acad Dental Mater 2016;32:43-53.
- 7. Barrère F, van Blitterswijk CA, De GK. Bone regeneration: molecular and cellular interactions with calcium phosphate ceramics. Int J Nanomed 2006;1:317-32.
- Davies JE. Bone bonding at natural and biomaterial surfaces. Biomaterials 2007;28:5058-67.
- Wopenka B, Pasteris JD. A mineralogical perspective on the apatite in bone. Mater Sci Engin C 2005;25:131-43.
- Friedman CD, Costantino PD, Takagi S, Chow LC. BoneSource hydroxyapatite cement: a novel biomaterial for craniofacial skeletal tissue engineering and reconstruction. J Biomed Mater Res 1998;43:428-32.
- Bohner M, Lemaitre J. Can bioactivity be tested in vitro with SBF solution? Biomaterials 2009;30:2175-9.
- Samavedi S, Whittington AR, Goldstein AS. Calcium phosphate ceramics in bone tissue engineering: a review of properties and their influence on cell behavior. Acta Biomater 2013;9:8037-45.
- Chavan PN, Bahir MM, Mene RU, et al. Study of nanobiomaterial hydroxyapatite in simulated body fluid: Formation and growth of apatite. Mater Sci Engin: B 2010;168:224-30.
- Großardt C, Ewald A, Grover LM, et al. Passive and active in vitro resorption of calcium and magnesium phosphate cements by osteoclastic cells. Tissue Engin Part A 2010;16:3687-95.
- 15. Zhou H, Lee J. Nanoscale hydroxyapatite particles for bone tissue engineering. Acta Biomater 2011;7:2769-81.
- 16. Kubasiewicz-Ross P, Hadzik J, Seeliger J, et al. New nano-hydroxyapatite in bone defect regeneration: A histological study in rats. Ann Anat [Anatomischer Anzeiger]: Official Organ Anatomische Gesellschaft 2017;213:83-90.

- de Freitas Silva L, de Carvalho Reis ENR, Barbara TA, et al. Assessment of bone repair in critical-size defect in the calvarium of rats after the implantation of tricalcium phosphate beta (beta-TCP). Acta Histochem 2017;119:624-31.
- Rolvien T, Barvencik F, Klatte TO, et al. β-TCP bone substitutes in tibial plateau depression fractures. Knee 2017;24:1138-45.
- Lindgren C, Mordenfeld A, Johansson CB, Hallman M. A 3-year clinical follow-up of implants placed in two different biomaterials used for sinus augmentation. Int J Oral Maxillofac Implants 2012;27:1151-62.
- Delécrin J, Takahashi S, Gouin F, Passuti N. A synthetic porous ceramic as a bone graft substitute in the surgical management of scoliosis: a prospective, randomized study. Spine 2000;25: 563-9.
- van Dijk LA, Barbieri D, Yuan H, DeBruijn J. Efficacy of biphasic calcium phosphate with submicron-scale surface topography as autograft extender in a rabbit posterolateral lumbar spine fusion model. Spine J 2018;18:S216-7.
- 22. Huang M-S, Wu H-D, Teng N-C, et al. In vivo evaluation of poorly crystalline hydroxyapatite-based biphasic calcium phosphate bone substitutes for treating dental bony defects. J Dental Sci 2010;5:100-8.
- 23. Fellah BH, Gauthier O, Weiss P, et al. Osteogenicity of biphasic calcium phosphate ceramics and bone autograft in a goat model. Biomaterials 2008;29:1177-88.
- 24. Hench LL, Polak JM. Third-Generation Biomedical Materials. Science 2002;295:1014-7.
- 25. Rahaman MN, Day DE, Bal BS, et al. Bioactive glass in tissue engineering. Acta Biomater 2011;7:2355-73.
- GonzÁLez-Carrasco JL. Metals as bone repair materials. In: Planell JA, Best SM, Lacroix D, Merolli A, eds. Bone Repair Biomaterials. Woodhead Publishing; 2009. pp 154-193.
- Sivakumar M, Kumar Dhanadurai KS, Rajeswari S, Thulasiraman V. Failures in stainless steel orthopaedic implant devices: A survey. J Mater Sci Lett 1995;14:351-4.
- Xu W, Yu F, Yang L, et al. Accelerated corrosion of 316L stainless steel in simulated body fluids in the presence of H2O2 and albumin. Mater Sci Engin: C 2018;92:11-9.
- 29. Manam NS, Harun WSW, Shri DNA, et al. Study of corrosion in biocompatible

metals for implants: A review. J Alloys Compound 2017;701:698-715.

- Chen Q, Thouas GA. Metallic implant biomaterials. Mater Sci Engin: R: Reports 2015;87:1-57.
- Stohs SJ, Bagchi D. Oxidative mechanisms in the toxicity of metal ions. Free Rad Biol Med 1995;18:321-36.
- 32. Thyssen JP. Nickel and cobalt allergy before and after nickel regulation-evaluation of a public health intervention. Contact Dermatitis 2011;65:1-68.
- 33. Chan BP, Hui TY, Wong MY, et al. Mesenchymal stem cell-encapsulated collagen microspheres for bone tissue engineering. Tissue Engin Part C Methods 2010;16:225-35.
- Zhang D, Wu X, Chen J, Lin K. The development of collagen based composite scaffolds for bone regeneration. Bioact Mater 2018;3:129-38.
- Omenetto FG, Kaplan DL. New opportunities for an ancient material. Science 2010;329:528-31.
- Kirker-Head C, Karageorgiou V, Hofmann S, et al. BMP-silk composite matrices heal critically sized femoral defects. Bone 2007;41:247-55.
- 37. Park SY, Ki CS, Park YH, et al. Electrospun silk fibroin scaffolds with macropores for bone regeneration: an in vitro and in vivo study. Tissue Engin Part A 2010;16:1271-9.
- Mieszawska AJ, Llamas JG, Vaiana CA, et al. Clay enriched silk biomaterials for bone formation. Acta Biomater 2011;7:3036-41.
- Wang Y, Rudym DD, Walsh A, et al. In vivo degradation of three-dimensional silk fibroin scaffolds. Biomaterials 2008;29:3415-28.
- 40. Bhattacharjee P, Kundu B, Naskar D, et al. Silk scaffolds in bone tissue engineering: An overview. Acta Biomater 2017;63:1-17.
- Farah S, Anderson DG, Langer R. Physical and mechanical properties of PLA, and their functions in widespread applications-A comprehensive review. Adv Drug Deliv Rev 2016;107:367-92.
- 42. Ramot Y, Haim-Zada M, Domb AJ, Nyska A. Biocompatibility and safety of PLA and its copolymers. Adv Drug Deliv Rev 2016;107:153-62.
- 43. Schrumpf MA, Lee AT, Weiland AJ. Foreign-body reaction and osteolysis induced by an intraosseous poly-l-lactic acid suture anchor in the wrist: case report. J Hand Surg 2011;36:1769-73.
- 44. Nusselt T, Freche S, Klinger HM, Baums MH. Intraosseous foreign body granuloma in rotator cuff repair with bioabsorbable suture anchor. Archiv Orthopaed Trauma Surg





2010;130:1037-40.

- 45. Murariu M, Dubois P. PLA composites: From production to properties. Adv Drug Deliv Rev 2016;107:17-46.
- 46. Coenen AMJ, Bernaerts KV, et al. Elastic materials for tissue engineering applications: Natural, synthetic, and hybrid polymers. Acta Biomater 2018;79:60-82.
- 47. Tatara AM, Mikos AG. Tissue Engineering in Orthopaedics. J Bone Joint Surg Am 2016;98:1132-9.
- Li X, Feng Q, Liu X, et al. Collagenbased implants reinforced by chitin fibres in a goat shank bone defect model. Biomaterials 2006;27:1917-23.
- 49. Chen L, Hu J, Ran J, et al. Preparation and evaluation of collagen-silk fibroin/hydroxyapatite nanocomposites for bone tissue engineering. Int J Biol Macromol 2014;65:1-7.
- 50. Zhang C, Zhu J, Zhang T, et al. Small intestinal submucosa/polymethyl methacrylate composite bone cement for vertebral repair. Mater Design 2018;154:254-65.
- 51. Yu W, Li R, Long J, et al. Use of a three-dimensional printed polylactidecoglycolide/tricalcium phosphate composite scaffold incorporating magnesium powder to enhance bone defect repair in rabbits. J Orthopaedic Transl 2018; [in press].
- 52. Coleman JN, Khan U, Blau WJ, Gun'Ko YK. Small but strong: A review of the mechanical properties of carbon nanotube–polymer composites. Carbon 2006;44:1624-52.
- 53. Ogihara N, Usui Y, Aoki K, et al. Biocompatibility and bone tissue compatibility of alumina ceramics reinforced with carbon nanotubes. Nanomedicine 2012;7:981-93.
- Abarrategi A, Gutiérrez MC, Moreno-Vicente C, et al. Multiwall carbon nanotube scaffolds for tissue engineering purposes. Biomaterials 2008;29:94-102.
- 55. Mukherjee S, Nandi SK, Kundu B, et al. Enhanced bone regeneration with carbon nanotube reinforced hydroxyapatite in animal model. J Mechan Behav Biomed Mater 2016;60:243-55.
- 56. Holzinger M, Vostrowsky O, Hirsch A, et al. Sidewall functionalization of carbon nanotubes. Angewandte Chemie Int Ed 2001;40:4002-5.
- 57. Venkatesan J, Ryu B, Sudha PN, Kim S-K. Preparation and characterization of chitosan-carbon nanotube scaffolds for bone tissue engineering. Int J Biol Macromol 2012;50:393-402.
- 58. Li X, Zhao T, Sun L, et al. The applications of conductive nanomaterials in

the biomedical field. J Biomed Mater Res Part A 2016;104:320-37.

- 59. Nayak TR, Jian L, Phua LC, et al. Thin films of functionalized multiwalled carbon nanotubes as suitable scaffold materials for stem cells proliferation and bone formation. ACS Nano 2010;4:7717-25.
- Stoller MD, Park S, Zhu Y, et al. Graphene-based ultracapacitors. Nano Lett 2008;8:3498-502.
- Lee C, Wei X, Kysar JW, Hone J. Measurement of the elastic properties and intrinsic strength of monolayer graphene. Science 2008;321:385-8.
- Nayak TR, Andersen H, Makam VS, et al. Graphene for controlled and accelerated osteogenic differentiation of human mesenchymal stem cells. ACS Nano 2011;5:4670-8.
- Kenry, Lee WC, Loh KP, Lim CT. When stem cells meet graphene: Opportunities and challenges in regenerative medicine. Biomaterials 2018;155: 236-50.
- Buwalda SJ, Boere KWM, Dijkstra PJ, et al. Hydrogels in a historical perspective: From simple networks to smart materials. J Control Release 2014;190: 254-73.
- Ferreira NN, Ferreira LMB, Cardoso VMO, et al. Recent advances in smart hydrogels for biomedical applications: From self-assembly to functional approaches. Eur Polymer J 2018;99: 117-33.
- 66. Bai X, Gao M, Syed S, et al. Bioactive hydrogels for bone regeneration. Bioact Mater 2018;3:401-17.
- 67. Ghosal A, Vashist A, Tiwari S, et al. Chapter 11 - Hydrogels: Smart nanomaterials for biomedical applications. In: Mohan Bhagyaraj S, Oluwafemi OS, Kalarikkal N, Thomas S, eds. Synthesis of inorganic nanomaterials. Woodhead Publishing; 2018. pp 283-292.
- Papathanasiou KE, Turhanen P, Brückner SI, et al. Smart, programmable and responsive injectable hydrogels for controlled release of cargo osteoporosis drugs. Scient Rep 2017;7:4743.
- 69. Ni P, Ding Q, Fan M, et al. Injectable thermosensitive PEG–PCL–PEG hydrogel/acellular bone matrix composite for bone regeneration in cranial defects. Biomaterials 2014;35:236-48.
- Uto K, Tsui JH, DeForest CA, Kim D-H. Dynamically tunable cell culture platforms for tissue engineering and mechanobiology. Progress Polymer Sci 2017;65:53-82.
- 71. Yang C, Tibbitt MW, Basta L, Anseth KS. Mechanical memory and dosing

influence stem cell fate. Nat Mater 2014;13:645-52.

- 72. Samal SK, Dash M, Dubruel P, Van Vlierberghe S. 8 - Smart polymer hydrogels: properties, synthesis and applications. In: Aguilar MR, San Román J, eds. Smart Polymers and their Applications. Woodhead Publishing; 2014. pp 237-270.
- 73. Mouriño V, Cattalini JP, Boccaccini AR. Metallic ions as therapeutic agents in tissue engineering scaffolds: an overview of their biological applications and strategies for new developments. J Royal Soc Interface 2012;9:401-19.
- Wang W, Yeung KWK. Bone grafts and biomaterials substitutes for bone defect repair: A review. Bioact Mater 2017;2:224-47.
- 75. Coulombe J, Faure H, Robin B, Ruat M. In vitro effects of strontium ranelate on the extracellular calcium-sensing receptor. Biochem Biophys Res Commun 2004;323:1184-90.
- Baron R, Tsouderos Y. In vitro effects of S12911-2 on osteoclast function and bone marrow macrophage differentiation. Eur J Pharmacol 2002;450:11-7.
- 77. Sangeetha K, Ashok M, Girija EK, et al. Strontium and ciprofloxacin modified hydroxyapatites as functional grafts for bone prostheses. Ceramics Int 2018;44:13782-9.
- Prabha RD, Nair BP, Ditzel N, et al. Strontium functionalized scaffold for bone tissue engineering. Mater Sci Engin: C 2019;94:509-15.
- Zhao D, Witte F, Lu F, et al. Current status on clinical applications of magnesium-based orthopaedic implants: A review from clinical translational perspective. Biomaterials 2017;112:287-302.
- Staiger MP, Pietak AM, Huadmai J, Dias G. Magnesium and its alloys as orthopedic biomaterials: A review. Biomaterials 2006;27:1728-34.
- Revell PA, Damien E, Zhang XS, et al. The effect of magnesium ions on bone bonding to hydroxyapatite coating on titanium alloy implants. Key Engin Mater 2004;447-50.
- Zreiqat H, Howlett CR, Zannettino A, et al. Mechanisms of magnesium-stimulated adhesion of osteoblastic cells to commonly used orthopaedic implants. J Biomed Mater Res 2002;62:175-84.
- 83. Bondarenko A, Angrisani N, Meyer-Lindenberg A, et al. Magnesium-based bone implants: Immunohistochemical analysis of peri-implant osteogenesis by evaluation of osteopontin and osteocalcin expression. J Biomed Mater Res



Review

Part A 2014;102:1449-57.

- Saris N-EL, Mervaala E, Karppanen H, et al. Magnesium: An update on physiological, clinical and analytical aspects. Clin Chim Acta 2000;294:1-26.
- 85. Bose S, Tarafder S, Bandyopadhyay A. Effect of chemistry on osteogenesis and angiogenesis towards bone tissue engineering using 3D printed scaffolds. Ann Biomed Engin 2017;45:261-72.
- Bhattacharjee P, Begam H, Chanda A, Nandi SK. Animal trial on zinc doped hydroxyapatite: A case study. J Asian Ceramic Soc 2014;2:44-51.
- Eltohamy M, Kundu B, Moon J, et al. Anti-bacterial zinc-doped calcium silicate cements: Bone filler. Ceramics Int 2018;44:13031-8.
- Lusvardi G, Malavasi G, Menabue L, et al. Properties of zinc releasing surfaces for clinical applications. J Biomater Appl 2008;22:505-26.
- Zhang D, Wong CS, Wen C, Li Y. Cellular responses of osteoblast-like cells to 17 elemental metals. J Biomed Mater Res Part A 2017;105:148-58.
- Holmes RE. Bone regeneration within a coralline hydroxyapatite implant. Plastic Reconstr Surg 1979;63:626-33.
- 91. Pountos I, Giannoudis PV. Is there a role of coral bone substitutes in bone repair? Injury 2016;47:2606-13.
- Ripamonti U. The morphogenesis of bone in replicas of porous hydroxyapatite obtained from conversion of calcium carbonate exoskeletons of coral. J Bone Joint Surg Am 1991;73:692-703.
- 93. Cong TT, Gargiulo C, Thao HD, et al. Culture and differentiation of osteoblasts on coral scaffold from human bone marrow mesenchymal stem cells. Cell Tissue Bank 2011;12:247-61.
- Alsalihi KA, Samsudin AR. Bone marrow mesenchymal stem cells differentiation and proliferation on the surface of coral implant. Med J Malaysia 2004;59:45-6.
- 95. Byrd HS, Hobar PC, Shewmake K. Augmentation of the craniofacial skeleton with porous hydroxyapatite granules. Plastic Reconstr Surg 1993;91:15-22; discussion 23-16.
- 96. Bizette C, Raul JS, Orhan B, et al. Results of cervical interbody fusion with coral grafts. Neuro-Chirurgie 1999;45:4-14.
- Ramzi N, Ribeiro-Vaz G, Fomekong E, et al. Long term outcome of anterior cervical discectomy and fusion using coral grafts. Acta Neurochir 2008;150: 1249-56.
- 98. Vuola J, Böhling T, Kinnunen J, et al.

Natural coral as bone-defect-filling material. J Biomed Mater Res Part B Appl Biomater 2000;51:117-22.

- 99. Wu YC, Lee TM, Chiu KH, et al. A comparative study of the physical and mechanical properties of three natural corals based on the criteria for bone-tissue engineering scaffolds. J Mater Sci Mater Med 2009;20:1273-80.
- 100. Terada M, Izumi K, Ohnuki H, et al. Construction and characterization of a tissue-engineered oral mucosa equivalent based on a chitosan-fish scale collagen composite. J Biomed Mater Res Part B: Appl Biomater 2012;100B:1792-802.
- 101. Capati MLF, Nakazono A, Yamamoto K, et al. Fish Collagen promotes the expression of genes related to osteoblastic activity. Int J Polymer Sci 2016;1-7.
- 102. Matsumoto R, Uemura T, Xu Z, et al. Rapid oriented fibril formation of fish scale collagen facilitates early osteoblastic differentiation of human mesenchymal stem cells. J Biomed Mater Res Part A 2015;103:2531-9.
- 103. Liu R, Holck AL, Yang E, et al. Tropomyosin from tilapia (Oreochromis mossambicus) as an allergen. Clin Experiment Allergy 2013;43:365-77.
- 104. Li X, Liu H, Niu X, et al. The use of carbon nanotubes to induce osteogenic differentiation of human adiposederived MSCs invitro and ectopic bone formation invivo. Biomaterials 2012;33:4818-27.
- 105. Li X, van Blitterswijk CA, Feng Q, et al. The effect of calcium phosphate microstructure on bone-related cells in vitro. Biomaterials 2008;29:3306-16.
- 106. Rao SH, Harini B, Shadamarshan RPK, et al. Natural and synthetic polymers/bioceramics/bioactive compounds-mediated cell signalling in bone tissue engineering. Int J Biol Macromol 2018;110:88-96.
- 107. Jung S-R, Song N-J, Yang DK, et al. Silk proteins stimulate osteoblast differentiation by suppressing the Notch signaling pathway in mesenchymal stem cells. Nutr Res 2013;33:162-70.
- 108. Chen Z, Wu C, Gu W, et al. Osteogenic differentiation of bone marrow MSCs by β -tricalcium phosphate stimulating macrophages via BMP2 signalling pathway. Biomaterials 2014;35:1507-18.
- 109. Chen Z, Wu C, Yuen J, et al. Influence of osteocytes in the *in vitro* and *in vivo* β -tricalcium phosphate-stimulated osteogenesis. J Biomed Mater Res Part A 2014;102:2813-23.

[BioMaterials Advances 2019; 2:63]

- 110. Zhang X, Zu H, Zhao D, et al. Ion channel functional protein kinase TRPM7 regulates Mg ions to promote the osteoinduction of human osteoblast via PI3K pathway: In vitro simulation of the bone-repairing effect of Mgbased alloy implant. Acta Biomater 2017;63:369-82.
- 111. Liu H, Peng H, Wu Y, et al. The promotion of bone regeneration by nanofibrous hydroxyapatite/chitosan scaffolds by effects on integrin-BMP/Smad signaling pathway in BMSCs. Biomaterials 2013;34:4404-17.
- 112. Lin Y, Huang Y, He J, et al. Role of Hedgehog-Gli1 signaling in the enhanced proliferation and differentiation of MG63 cells enabled by hierarchical micro-/nanotextured topography. Int J Nanomed 2017;12:3267-80.
- 113. Pan H, Xie Y, Zhang Z, et al. Hierarchical macropore/nano surface regulates stem cell fate through a ROCK-related signaling pathway. Rsc Advances 2017;7:8521-32.
- 114. Wang W, Liu Q, Zhang Y, Zhao L. Involvement of ILK/ERK1/2 and ILK/p38 pathways in mediating the enhanced osteoblast differentiation by micro/nanotopography. Acta Biomater 2014;10:3705-15.
- 115. Liu N, Zhou M, Zhang Q, et al. Stiffness regulates the proliferation and osteogenic/odontogenic differentiation of human dental pulp stem cells via the WNT signalling pathway. Cell Prolif 2018;51:e12435.
- 116. Yuan H, Zhou Y, Lee MS, et al. A newly identified mechanism involved in regulation of human mesenchymal stem cells by fibrous substrate stiffness. Acta Biomater 2016;42:247-57.
- 117. Zhang K, Fan Y, Dunne N, Li X. Effect of microporosity on scaffolds for bone tissue engineering. Regener Biomater 2018;5:115-24.
- 118. Li X, Liu H, Niu X, et al. The use of carbon nanotubes to induce osteogenic differentiation of human adiposederived MSCs in vitro and ectopic bone formation in vivo. Biomaterials 2012;33:4818-27.
- 119. Ren X, Tuo Q, Tian K, et al. Enhancement of osteogenesis using a novel porous hydroxyapatite scaffold in vivo and vitro. Ceramics Int 2018;44:21656-65.
- 120. Du M, Zhu T, Duan X, et al. Acellular dermal matrix loading with bFGF achieves similar acceleration of bone regeneration to BMP-2 via differential effects on recruitment, proliferation and sustained osteodifferentiation of mesenchymal stem cells. Mater Sci



Review

Engin: C 2017;70:62-70.

- 121. Shih YR, Phadke A, Yamaguchi T, et al. Synthetic bone mimetic matrixmediated in situ bone tissue formation through host cell recruitment. Acta Biomater 2015;19:1-9.
- 122. Phadke A, Hwang Y, Kim SH, Kim SH, Yamaguchi T, Masuda K, et al. Effect of scaffold microarchitecture on osteogenic differentiation of human mesenchymal stem cells. Eur Cells Mater 2013;25:114-28; discussion 128-119.
- 123. Song G, Habibovic P, Bao C, et al. The homing of bone marrow MSCs to nonosseous sites for ectopic bone formation induced by osteoinductive calcium phosphate. Biomaterials 2013;34:2167-76.
- 124. Li Y, Guo Y, Ge J, et al. In situ silica nanoparticles-reinforced biodegradable poly(citrate-siloxane) hybrid elastomers with multifunctional properties for simultaneous bioimaging and bone tissue regeneration. Appl Mater Today 2018;10:153-63.
- 125. Quinlan E, Partap S, Azevedo MM, et al. Hypoxia-mimicking bioactive glass/collagen glycosaminoglycan composite scaffolds to enhance angiogenesis and bone repair. Biomaterials 2015;52:358-66.
- 126. Kesireddy V, Kasper FK. Approaches for building bioactive elements into synthetic scaffolds for bone tissue engineering. J Mater Chem B 2016;4: 6773-86.

- 127. Costa F, Carvalho IF, Montelaro RC, et al. Covalent immobilization of antimicrobial peptides (AMPs) onto biomaterial surfaces. Acta Biomater 2011;7:1431-40.
- 128. Visser R, Rico-Llanos GA, Pulkkinen H, Becerra J. Peptides for bone tissue engineering. J Control Release 2016;244: 122-35.
- 129. Zhao X, Han Y, Li J, et al. BMP-2 immobilized PLGA/hydroxyapatite fibrous scaffold via polydopamine stimulates osteoblast growth. Mater Sci Engin: C 2017;78:658-66.
- 130. Kuttappan S, Mathew D, Jo J-i, et al. Dual release of growth factor from nanocomposite fibrous scaffold promotes vascularisation and bone regeneration in rat critical sized calvarial defect. Acta Biomater 2018;78:36-47.
- 131. Chen Z, Yuen J, Crawford R, et al. The effect of osteoimmunomodulation on the osteogenic effects of cobalt incorporated β-tricalcium phosphate. Biomaterials 2015;61:126-38.
- 132. Pajarinen J, Lin T, Gibon E, et al. Mesenchymal stem cell-macrophage crosstalk and bone healing. Biomaterials 2018 [in press].
- 133. Chen Z, Klein T, Murray RZ, et al. Osteoimmunomodulation for the development of advanced bone biomaterials. Mater Today 2016;19:304-21.
- 134. Hotchkiss KM, Clark NM, Olivares-Navarrete R. Macrophage response to hydrophilic biomaterials regulates MSC recruitment and T-helper cell

populations. 2018;182:202-15.

Biomaterials

- 135. Miao S, Castro N, Nowicki M, et al. 4D printing of polymeric materials for tissue and organ regeneration. Mater Today 2017;20:577-91.
- 136. Blokhuis TJ, Arts JJ. Bioactive and osteoinductive bone graft substitutes: definitions, facts and myths. Injury 2011;42:26-9.
- 137. Lienau J, Schmidt-Bleek K, Peters A, et al. Differential regulation of blood vessel formation between standard and delayed bone healing. J Orthopaed Res 2009;27;1133-40.
- 138. Zhou Y, Shi M, Jones JR, et al. Strategies to direct vascularisation using mesoporous bioactive glassbased biomaterials for bone regeneration. Int Mater Rev 2017;62:392-414.
- 139. Kang Y, Kim S, Fahrenholtz M, et al. Osteogenic and angiogenic potentials of monocultured and co-cultured human-bone-marrow-derived mesenchymal stem cells and human-umbilical-vein endothelial cells on threedimensional porous beta-tricalcium phosphate scaffold. Acta Biomater 2013;9:4906-15.
- 140. Hadjipanayi E, Brown RA, Mudera V, et al. Controlling physiological angiogenesis by hypoxia-induced signaling. J Control Release 2010;146:309-17.
- 141. Laschke MW, Menger MD. Prevascularization in tissue engineering: Current concepts and future directions. Biotechnol Adv 2016;34:112-21.